Rapid and Sensitive Detection of Severe Acute Respiratory Syndrome Coronavirus by Rolling Circle Amplification

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The severe acute respiratory syndrome (SARS) epidemic of 2003 was responsible for 774 deaths and caused significant economic damage worldwide. Since July 2003, a number of SARS cases have occurred in China, raising the possibility of future epidemics. We describe here a rapid, sensitive, and highly efficient assay for the detection of SARS coronavirus (SARS-CoV) in cultured material and a small number (n = 7) of clinical samples. Using rolling circle amplification (RCA), we were able to achieve sensitive detection levels of SARS-CoV RNA in both solid and liquid phases. The main advantage of RCA is that it can be performed under isothermal conditions with minimal reagents and avoids the generation of false-positive results, a problem that is frequently encountered in PCR-based assays. Furthermore, the RCA technology provides a faster, more sensitive, and economical option to currently available PCR-based methods.

Severe acute respiratory syndrome (SARS) is an emerging disease caused by the novel SARS coronavirus (SARS-CoV) (2, 4, 5, 14). By the end of the SARS epidemic in July 2003, a total of 8,096 SARS cases had been reported from 30 countries, with 774 deaths. Whether future outbreaks of SARS will occur is unknown at present. However, given the recent SARS cases in southern China arising from an unknown source and a number of laboratory-related infections (12), it is important to be prepared for such a possibility. In the absence of a SARS-CoV vaccine or antiviral drugs, the use of strict infection control policies and early diagnosis with rapid, sensitive, and highly specific laboratory methods are essential for the early management of SARS-CoV infection.

Apart from epidemiological linkages, the clinical and radiographic features of the disease are not SARS specific, identifying a need for specific laboratory tests that can confirm SARS-CoV infection early in the course of the illness. Detection of SARS-CoV-specific antibodies is a sensitive and specific but is not possible at clinical presentation (6, 14).

Detection of SARS-CoV by reverse transcription-PCR (RT-PCR) in clinical specimens allows diagnosis in the early stage of the disease. However, in contrast to many other acute respiratory infections, only low levels of SARS-CoV are thought to be present during the early symptomatic phase of infection. On the basis of the results of first-generation RT-PCR assays, SARS-CoV RNA can be detected with a sensitivity of only ca. 30 to 50% in a single respiratory specimen. A higher sensitivity can be achieved if serial samples are collected, particularly during the second week of illness when maximal virus shedding occurs (13, 14). The type of clinical sample (e.g., nasopharyngeal aspirate, throat swabs, stool samples, urine, etc.) also affects the sensitivity of RT-PCR (21).

Recently, the utility of circularizable oligonucleotides, or "padlock probes," has been demonstrated for the detection of target nucleic acid sequences; this approach shows greater sensitivity than conventional PCR (3, 8, 16). Upon hybridization to a target DNA or RNA sequence, the two ends of the probe become juxtaposed and can be joined by DNA ligase (Fig. 1). The circularized DNA probe then creates an effective template for an exponential, or hyperbranching, rolling-circle amplification (RCA) reaction (Fig. 1), catalyzed by a highly processive DNA polymerase with strand displacement activity. In isothermal conditions, hyperbranching RCA is capable of a 109-fold signal amplification of each circle within 90 min (8). The RCA technique is highly sensitive, and a circularized DNA probe bound to a single target template can be efficiently detected (3). The RCA assay also offers several advantages over other amplification techniques: the ligation requires Watson-Crick base pairing at both ends of the probe hybridize with perfect complementarity, not only permitting the detection of a single-nucleotide polymorphism but also preventing the nonspecific amplification generated by conventional PCR. Circularizable probes can be used for the recognition of both DNA and RNA templates, eliminating the need for RT and creating a uniform assay format for both RNA and DNA detection (11). Single-stranded DNA displaced by the DNA polymerase can be readily bound by primers, thus enabling the reaction to be performed under isothermal conditions and removing the need for a thermocycler. We describe here a simple, scalable assay using RCA technology that allows the rapid, sensitive, and efficient detection of cultured SARS virus in both liquid and solid phases and present preliminary results on a small number of clinical respiratory specimens.
A total of seven circularizable probes were designed targeting various regions of SARS-CoV (RNA and cDNA templates) (Table 1), carefully selected to ensure that probe binding was SARS-CoV-specific and not sharing similarity to other coronaviruses such as OC43 and 229E strains. The spacer regions were also designed to allow specific RCA primer binding and amplification of the probe-specific signal. To demonstrate a working molecular model and to allow optimization of assay conditions, artificial DNA, or RNA templates were synthesized (Table 1).

**Molecular model of RCA in the liquid phase.** Liquid-phase RCA was performed in two steps. The first was a ligation reaction, wherein the probe specifically binds the target template and becomes circularized by DNA ligase. Probe (20 pmol) was hybridized with 10^3 artificial single-stranded DNA molecules and circularized by incubation with 20 U of T4 DNA ligase (New England Biolabs) in 1× reaction buffer at 37°C for 30 min in a 20-μl reaction volume. Ligations using artificial RNA templates were performed by using 40 U of T4 DNA ligase with 1× reaction buffer containing 10 mM MgCl2 and 10 μM ATP in a total volume of 20 μl. Separate control reactions were also performed with human genomic DNA as a template. Five units of ampligase (a thermostable DNA ligase for double-stranded DNA [dsDNA]; Astral Scientific, Carrington, Australia) were used with human genomic DNA (1 μg) and circularizable probe (20 pmol). Genomic DNA was denatured at 95°C for 5 min, and a ligation step then performed at 45°C for 25 min.

After ligation and circularization of the probe, a second step was performed to amplify the circular probe by RCA using DNA polymerase with strand-displacing activity. Two DNA polymerases were chosen to perform the RCA: Vent (exo-) DNA polymerase and Bst DNA polymerase (New England Biolabs). RCA reactions were performed in a 50-μl volume containing Bst DNA polymerase (8 U) or Vent DNA polymerase (20 U), 400 μM deoxynucleoside triphosphate mix, 40 pmol of each RCA primer, and 5 μl of the original ligation mix. Circularized probe signals were amplified by incubation at 65°C for 90 min and visualized on a 1.5% agarose gel under UV illumination.

To optimize the reaction and further increase amplification efficiency, we modified the reaction mix by adding bacteriophage T4 Gene 32 protein (Roche Applied Science), a single-stranded DNA-binding protein. Gene 32 protein can destabilize dsDNA helices (10) and has been used to enhance PCR (7). During RCA reactions with 500 copies of artificial template, T4 gene 32 protein was added to the reaction in decreasing concentrations (5 μg to 0.5 ng/reaction). In separate experiments, 1 to 10% dimethyl sulfoxide (DMSO) was added to RCA reactions. DMSO can destabilize dsDNA helices while stabilizing polymerase-template complexes (20), thus further increasing the efficiency of the reaction.

**Molecular model of RCA in the solid phase.** Excess probe molecules carried over from the ligation step may bind RCA primers and form short dsDNA molecules, a process that may increase background signals and hamper quantitative analysis. To circumvent this, a solid-phase RCA assay was designed by using magnetic beads coated with oligo(dT) (Dynal mRNA DIRECT Kit, Dynal Biotech, Oslo, Norway) which bind mRNA molecules and provide a solid platform for the RCA reaction. Artificial RNA templates with a 3′-poly(A) tail were used to assess the sensitivity of this assay. Serially diluted artificial RNA templates were incubated with 40 μl of oligo(dT)-coated magnetic beads and 40 pmol of probe at room temperature for 10 min. The artificial RNA templates and probe complexed with beads were extracted by magnetic separation according to the manufacturer’s protocol. Probe bound to artificial RNA templates was circularized by incubation with 40 U of T4 DNA ligase in 1× reaction buffer containing 10 mM MgCl2 and 10 μM ATP at 37°C for 30 min. The RCA reaction was performed as described above in the liquid phase by using Bst DNA polymerase.

**Detection of SARS-CoV RNA target.** After the specificity and sensitivity of RCA was confirmed by molecular modeling, the assay was then tested on SARS-CoV RNA samples derived from cultured material and sputum samples of SARS patients. In solid-phase RCA, SARS-CoV RNA was extracted by incubation of specimens with 500 μl of lysis/binding buffer (supplied by the kit) for 10 min at room temperature, followed by the addition of oligo(dT)-coated magnetic beads (40 μl) to bind to RNA molecules. The RNA-bonded beads were washed four times with the wash buffer provided in the kit at room temperature and separated by using a magnet to remove possible inhibitors. Then 40 pmol of probe was added and mixed with bead-bonded viral RNA molecules. The ligation of probe and viral RNA was performed in by incubation with 40 U of T4 ligase in 1× reaction buffer (10 mM MgCl2 and 10 μM ATP) at 37°C for 30 min. After the ligation, a second washing step was carried out with the wash buffer provided with the kit to remove the unbound probe, and the circularized probe, viral RNA, and magnetic beads complex were separated by using a magnet. RCA reactions were then performed as described above.

**MATERIALS AND METHODS**

**Culture SARS-CoV and clinical specimens.** In cultured material, SARS CoV (HKU-39849) isolated from a patient with SARS CoV pneumonia was propagated in Vero cells in Dulbecco modified Eagle medium with 10% fetal calf serum. After single passage in Vero cells, viral RNA was extracted by QIAamp virus RNA minikit (Qiagen) according to manufacturer’s protocol. SARS-CoV RNA was kindly provided by J. S. M. Peiris at the Department of Microbiology, University of Hong Kong and Queen Mary Hospital, Hong Kong. The sputum samples from seven SARS patients (confirmed by RT-PCR) were collected at day 5 to 13 after the onset of illness. These sputum samples were collected by the National Institute of Virology Beijing, during April and May 2003 by the team of Yunle Hou.

**Circularizable probe design.** The oligonucleotide probes used in these experiments were ca. 80 to 90 nucleotides (nt) in length, consisting of two adjacent target complementary sequences (15 to 20 nt) with a spacer region (40 to 50 nt) to facilitate the loop formation and provide a template for RCA primer binding.
RESULTS

Circularizable probe design and amplification of probe signal using RCA. Seven circularizable probes were designed to target various RNA and cDNA regions of SARS-CoV (Table 1). To demonstrate a working molecular model, complementary artificial DNA or RNA templates were also synthesized. Initial testing of the seven probes and their corresponding artificial templates indicated that six of seven probes readily bound to the template and probe signals could be easily detected (Fig. 2A). One probe did not generate a signal from its corresponding template, a finding that could be due to poor quality of the probe and/or secondary structure(s) in the template/probe. When human DNA samples were used as non-saturation template, no signals were observed, indicating highly specific binding for all six probes (Fig. 2B).

RCA in the liquid phase. To assess the sensitivity of the technique, RCA was performed on serial dilutions of the target template (artificial DNA and RNA templates), and a sensitivity of detection close to the single template level was achieved. However, the signal derived was relatively weak in many cases when the target template copy number was below 100 (Fig. 2C). Several strategies have been indicated to improve signal strength in RCA, including the addition of DMSO or T4 gene 32 protein (7, 20, 22) to destabilize dsDNA helices and stabilize polymerase-template complexes. Tests with 500 copies of artificial template and T4 gene 32 protein with decreasing concentrations from 5 μg to 0.5 ng/reaction indicated a minor enhancement of signal at a concentration of 0.5 ng (Fig. 2D).

In a separate experiment, 1 to 10% DMSO added to the RCA reaction by using 500 copies of artificial template gave no significant signal enhancement (data not shown). More tests are required to validate optimal conditions for use of these enhancers to maximize signal strength. In addition to weak signal strength at low copy numbers, unbound probe molecules carried over from the ligation mix sometimes resulted in smear-like background during the liquid-phase RCA assay. The presence of background signal makes liquid-phase RCA detection of SARS-CoV less suitable for possible quantitative applications.

RCA in the solid phase. To circumvent these problems and prevent the formation of background signals, we developed a variant “solid-phase” RCA assay using magnetic beads coated with oligo(dT). The poly(A) signal located at the 3' end of the cDNA transcripts was ligated to the magnetic beads, facilitating the rapid isolation of RNA and further providing a solid-phase medium for the RCA reaction. A molecular model was also established by using artificial RNA templates with poly(A) sequences located at the 3' end. Serial dilutions of template used in RCA proved that the solid-phase method could detect close to a single artificial copy with a strong signal (Fig. 2E).

Detection of SARS-CoV RNA by RCA. The molecular models we developed demonstrated the sensitivity of RCA for SARS-CoV detection, particularly in the solid phase. Testing the SARS-CoV RNA samples derived from cultured material with known copy numbers clearly indicated that as few as five copies of SARS viral genome (defined by using a RealArt HPA Coronavirus kit; RealArt, Hamburg, Germany) could be easily detected by RCA in the solid phase (Fig. 3A). Testing of SARS-CoV from patient samples was also performed. In total,
all seven RNA samples derived from sputum of SARS-positive patients for SARS-CoV by RT-PCR showed a strong positive signal by RCA (Fig. 3B). Control samples of healthy individuals showed no signal in solid-phase RCA experiments.

**DISCUSSION**

This report describes an efficient method for the sensitive detection of SARS-CoV genome using RCA in both liquid and solid phases.

Given the occasional reappearance of SARS after the 2003 epidemic ended, the development of diagnostic assays for early, rapid, and reliable detection of SARS-CoV should remain a high priority. At present, the RT-PCR-based assays are the most widely used initial diagnostic tests. However, the sensitivity of SARS-CoV RT-PCR has been estimated to be as low as 30%, depending on the type of specimens tested and the timing of collection relative to the onset of symptoms (13, 14). One recent report has described an increase in sensitivity for the detection of the virus, benefiting earlier diagnosis (15). Despite the obtainable magnitude of amplification, these methods are time-consuming (e.g., RNA extraction, RT, and real-time PCR) and require a high-precision thermocycler, along with expensive reagents. In addition, cross-contamination and carryover, which lead to false-positive results, remains a continued problem. RCA offers a distinct advantage because it is based on the amplification of signal rather than target amplification.

The availability of ligases that accurately distinguish DNA
sequences has made oligonucleotide ligation assays increasingly popular in clinical diagnostics (1). DNA ligases have been isolated from a number of organisms and characterized for their ability to discriminate between matched and mismatched substrates (9, 17, 18, 19). The ATP-dependent T4 DNA ligase can join DNA oligonucleotides on RNA targets, although this reaction is quite inefficient at ATP concentrations commonly used for DNA-templated DNA ligation. Recently, however, reaction conditions have been defined that allow efficient RNA-template DNA ligation by the T4 DNA ligase (11), extending the use of padlock probes to RNA targets.

Although RCA has been shown to be highly sensitive and able to detect a single template (3, 8, 16), PCR remains the method of choice in both research and clinical diagnostics for sensitive detection of nucleic acids. However, in recognition of the promising features and considerable advantages of RCA and similar methods, padlock probes are becoming more widely adopted. PCR involves exponential target amplification, thereby increasing the risk of amplicon cross-contamination. In contrast, RCA specifically amplifies the probe signal avoiding the target template accumulation over time, minimizing the risk of contamination and making it more attractive for high-throughput analysis. Moreover, RCA can be performed in isothermal conditions, a factor that is useful where access to scientific equipment is minimal.

In the present study, we have successfully adopted the RCA technique for the detection of SARS-CoV from artificial and patient-derived material and have clearly demonstrated the advantages of this sensitive and specific diagnostic test. In molecular modeling with artificial DNA templates, we demonstrated that this technique is capable detecting down to the single template level. These results demonstrate a considerable potential of this technique in future clinical diagnostic testing of SARS-CoV, although the enhancement of signals in the liquid phase by DMSO and T4 gene 32 protein at low copy numbers of the template may require further optimization. In addition, the direct attachment to RNA molecules by circular probes can eliminate the RT step required by many RNA studies, providing a useful mechanism for the sensitive and accurate detection and identification of RNA sequences. The recent development of efficient RNA-templated DNA ligation (11), coupled with the powerful signal amplification generated by RCA down to the detection of a single circular probe (3), has facilitated the design of a sensitive assay for SARS-CoV RNA detection. More than 80% of the RNA template present can be recognized by DNA probes (11) and successfully ligated by T4 DNA ligase, making RCA detection of RNA template greater in sensitivity than conventional RT-PCR technology.

The relatively abundant unbound probe remaining in the ligation reaction during liquid-phase RCA can in some cases generate a background signal interfering with signal detection. This may be of concern, particularly in quantitative applications using fluorescently labeled primer at low copy numbers. However, the application of a solid-phase assay using oligo(dT)-coated magnetic beads provides promising solutions to this problem. This solid-phase format has the added advantage of rapid RNA extraction through the binding of oligo(dT) molecules on the beads, with poly(A) signals located at 3' end of the mRNAs. Hybridization between the target RNA and complementary probe binding region results in the formation of a helix with four turns (10 nt per turn, 20 nt long in each complementary region), allowing a ligation reaction to create a padlock structure. This interaction can withstand stringent washing, allowing the removal of unbound probe molecules. The solid-phase RCA assay demonstrated close to single artificial RNA template detection in molecular modeling with stronger signal, and five copies of SARS-CoV RNA were easily detectable. This clearly demonstrates the bright future of RCA in the diagnosis of SARS-CoV and indeed of other diseases. Testing conducted on patient samples further highlighted the usefulness of this assay as a future diagnostic test for SARS-CoV. It remains unclear why solid-phase RCA can generate a stronger signal than the liquid phase, but the reduction of background signal by solid-phase alone confirms it as an attractive option for ultrasensitive detection. Nonetheless, it is important to reiterate that we could successfully achieve low-copy detection in both solid- and liquid-phase RCA. Our data also showed that the RCA reaction is most efficient when catalyzed by thermostable strand-displacing enzymes such as Bst DNA polymerase. Additional work will be required to develop the ideal conditions for RCA in SARS-CoV detection, although the reaction sensitivity and signal strength already compares very favorably with PCR. Future studies are required for the development of a quantitative assay. Studies performed by Faruqi et al. (3) clearly demonstrated the quantitative potential of RCA, showing it to be capable of accurately detecting
numbers of circularizable probe bound to genomic DNA. We predict that the inherent simplicity of this method will allow RCA to be used in a wide variety of clinical applications.

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